

IN THE NAME OF GOD

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Research Communication

Aberrant Methylation of *APC* and *RAR β ₂* Genes in Breast Cancer Patients

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2014 ; in Egypt

Estimated Deaths

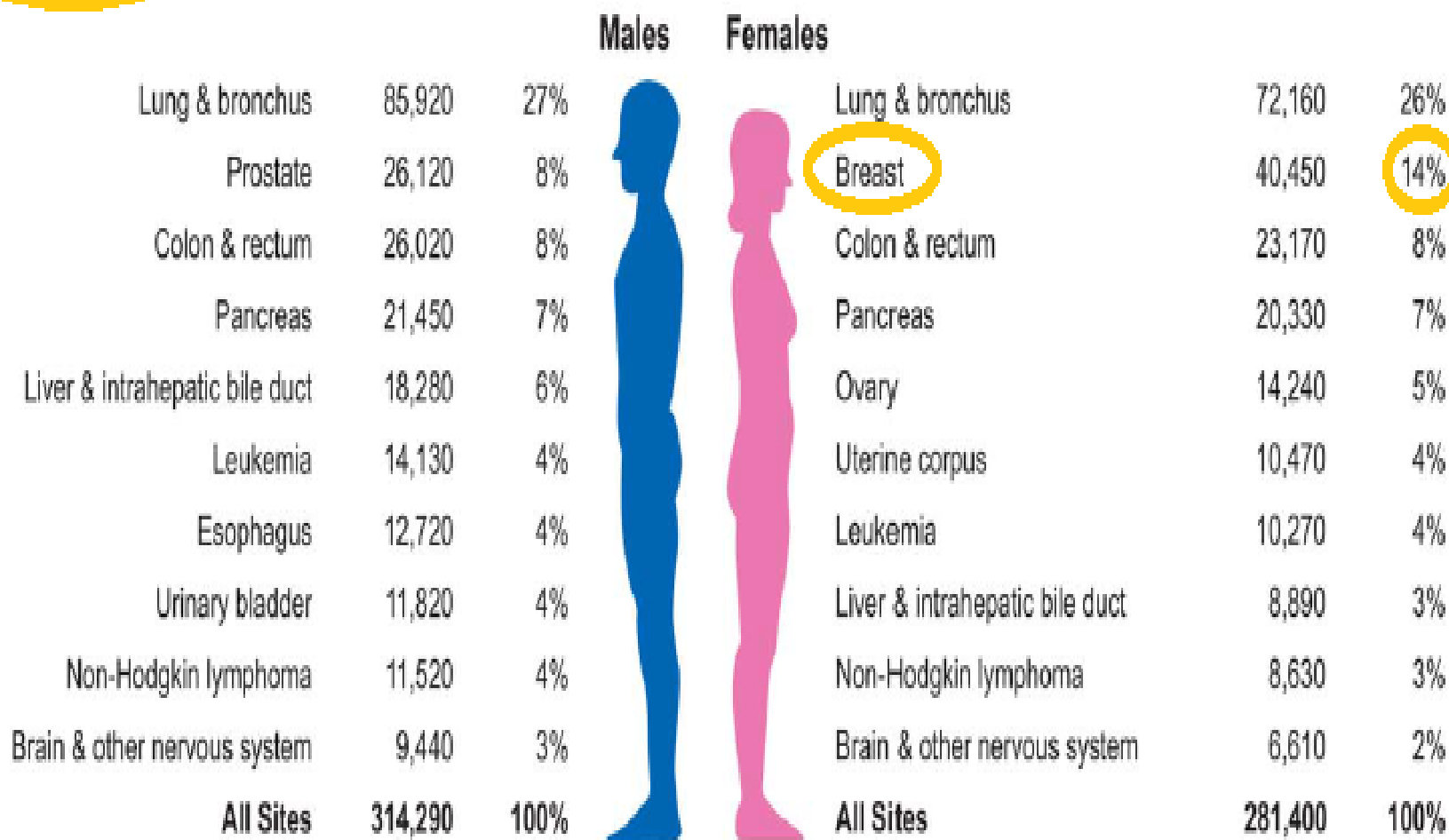


FIGURE 1. Ten Leading Cancer Types for the Estimated New Cancer Cases and Deaths by Sex, United States, 2016.

Estimates are rounded to the nearest 10 and cases exclude basal cell and squamous cell skin cancers and in situ carcinoma except urinary bladder.

○ Unmethylated

● Methylated



□ Samples :

- 121 malignant breast cancer (71/121 Triple negative)
- 79 benign Lesion
- 66 Healthy control

alternative mechanism for inactivation of tumor suppressor genes (5). Because gene hypermethylation has been found to be a common and early alteration in many tumor types (6), including breast (7), it has emerged as a promising target for detection of breast cancer. Several tumor suppressor genes have been found to be hypermethylated in breast cancer and unmethylated in normal cells, *e.g.*, *adenomatous polyposis coli* (APC) (8), and *retinoic acid receptors- β_2* (RAR- β_2) (9).

Detection of methylated DNA in the serum of cancer patients is quite intriguing and has been suggested to be a marker for early cancer development (10). Their concentration in cancer patients is higher than that from healthy people (11). Methylation specific-polymerase chain reaction (PCR) (MSP) a technique that can be used to detect hypermethylations as it can identify up to 1 methylated allele in 1000 unmethylated alleles, appropriate for the detection of neoplastic cells in a background of normal cells (12). It has been used in recent studies for the successful detection of cancer cell DNA in body fluids such as liver (13), lung (14), and head and neck cancer in serum (15), lung cancer in both sputum (16) and bronchial lavage (17), and bladder cancer in urine (18,19). Overall, these studies have demonstrated a high concordance between the epigenetic alterations found in primary tumor specimens and in body fluids, suggesting the potential utility of these alterations as surrogate molecular markers. Thus far, however,

teers. After obtaining informed consent from participants for the use of blood in this study, all of them were interviewed to elicit information regarding demographic characteristics; full clinical and pathological data were obtained from medical reports.

Collection and Processing of Samples and Assessment of Classical Tumor Markers

At the enrollment visit, 5 ml of blood was drawn into tube and then centrifuged at 1600g for 15 min at 15–20°C. The serum was stored at –80°C until processing for assessment of classical tumor markers and DNA extraction. Classical tumor markers (CEA and CA 15.3) were done by solid phase enzyme linked immunosorbent assay (Signosis, Sunnyvale).

Detection of Methylated RAR β_2 and APC Genes

This was accomplished in four steps. In the first step, DNA was extracted from serum samples. In the second step, DNA was treated by bisulfite. Third step, the gene was amplified by PCR. Finally, the fourth step included detection of the PCR products using agarose gel.

1. DNA isolation from serum samples

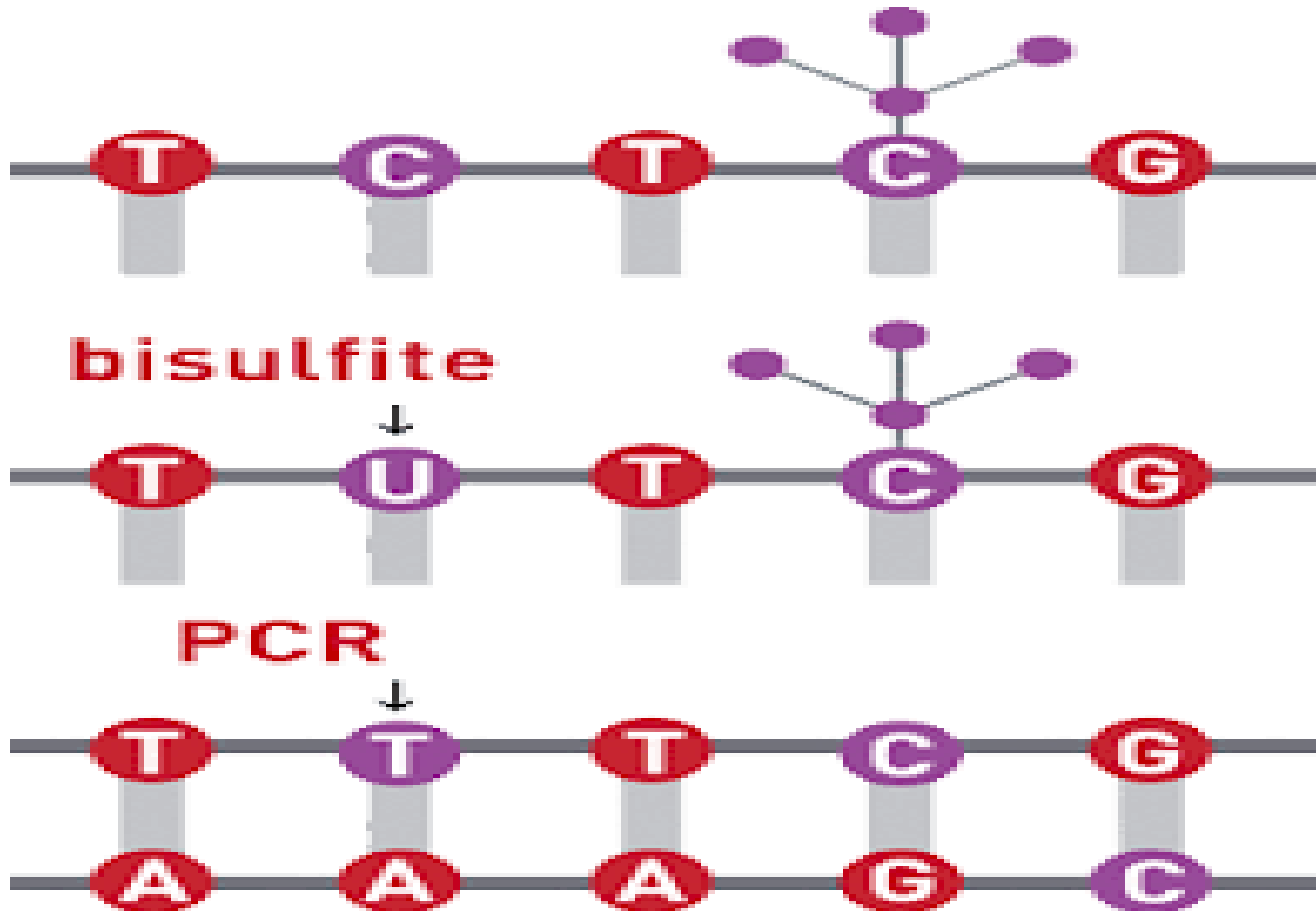
DNA extraction was done using QIAamp DNA Mini blood kit (QIAGEN, USA). In brief, 200 μ l serum was added with 20 μ l QIAGEN protease into microcentrifuge tube, samples were

○ Detection of Methylated **RARB2** and **APC** Genes

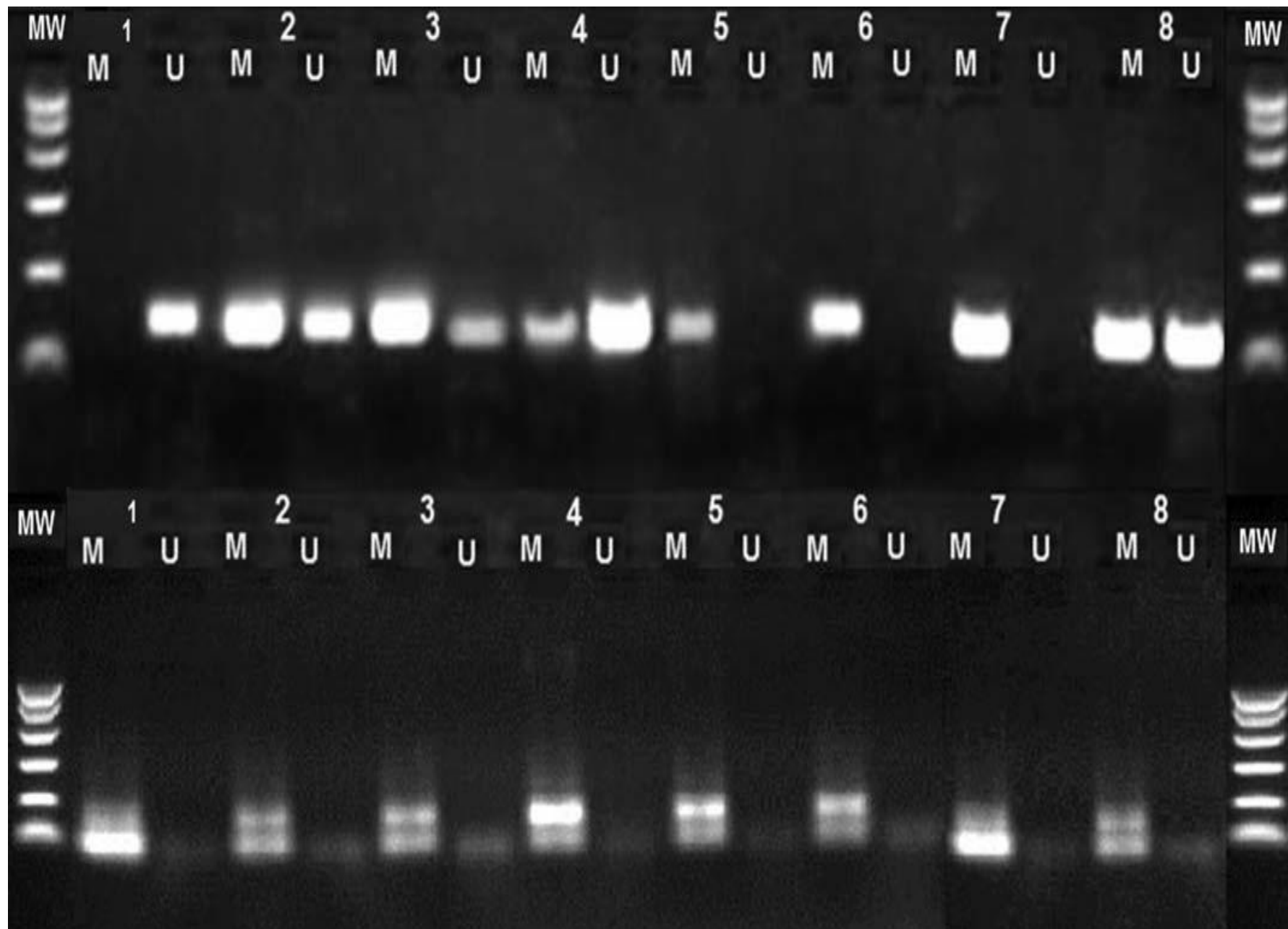
was accomplished in four steps:

1. DNA Extraction (by use of QIAamp DNA Mini blood Kit)
2. Treat by bisulfite
3. Amplified by PCR
4. Gel Electrophoresis

Sodium bisulfate treatment



- ❑ PCR condition in 50 µl volume of reaction
- Activation at 95°C for 5 min ; then 35cycles
- Denaturation at 94°C for 1 min
- Annealing 1 min ;
 - For APC at 53 °C
 - For RARB2 at 50 °C
 - For hMLH1 at 58 °C
- Extension at 72°C for 1 min
- Final extension at 72°C for 10 min



<i>Patient's characteristics</i>	<i>Cases (n = 121)</i>
Age (years)	
≤42	54 (44.6%)
>42	67 (45.4%)
Menopausal status	
Premenopausal	94 (77.7%)
Postmenopausal	27 (22.3%)
ER-status	
Negative	46 (38%)
Positive	75 (62%)
PgR-status	
Negative	61 (50.4%)
Positive	60 (49.6%)
HER-2/neu status	
Negative	45 (44.6%)
Positive	67 (55.4%)
Clinical stage	
Early stage	86 (71%)
Late stage	35 (29%)
Histological grade	
Low grade	89 (73.6%)
High grade	32 (26.4%)

TABLE 2

APC and RAR β_2 methylation pattern and positivity levels of tumor markers (CEA and CA15.3) among different investigated groups

<i>Investigated markers</i>	<i>Control (n = 66)</i>	<i>Benign (n = 76)</i>	<i>Malignant (n = 121)</i>
Methylated APC (n = 119)	0 (0%)	6 (7.8%)	113 (93.4%)
Homoplasmic methylation (n = 109)	0 (0%)	0 (0%)	109 (90.1)
Heteroplasmic methylation (n = 10)	0 (0%)	6 (7.8%)	4 (3.3%)
Methylated RAR β_2 (n = 127)	0 (0%)	11 (14.5%)	116 (95.9%)
Homoplasmic methylation (n = 113)	0 (0%)	9 (11.8%)	104 (86%)
Heteroplasmic methylation (n = 14)	0 (0%)	2 (2.6%)	12 (9.9%)
Positive CEA (>5 ng/ml)	0 (0%)	34 (44.7%)	63 (52.1%)
Positive CA15.3 (>15 ng/ml)	0 (0%)	20 (26.3%)	91 (75.2%)

<i>Patient's characteristics</i>	<i>Methylated RARβ_2</i>	<i>Methylated APC</i>	<i>Positive CEA</i>	<i>Positive CA15.3</i>
<i>Age (years)</i>				
≤ 42	51 (94.4%)	51 (94.4%)	33 (61.1%)	43 (79.6%)
> 42	65 (97%)	62 (92.5%)	30 (44.8)	48 (71.6%)
<i>Menopausal status</i>				
Premenopausal	91 (96.8%)	87 (92.6%)	52 (55.3%)	74 (78.7%)
Postmenopausal	25 (92.6%)	26 (96.3%)	11 (40.7%)	17 (63%)
<i>TNB</i>				
Cases with TNB	71 (100%)	67 (94.4%)	34 (47.9%)	53 (74.6%)
Cases with non-TNB	45 (90%)	46 (92.2%)	29 (58%)	38 (76%)

$$\chi^2 = 7.4, P = 0.007$$

	<i>Breast cancer</i>		<i>Early stage</i>		<i>Low grade</i>		<i>Triple negative breast cancer</i>	
<i>Investigated items</i>	<i>Sens.%</i>	<i>Spec.%</i>	<i>Sens.%</i>	<i>Spec.%</i>	<i>Sens.%</i>	<i>Spec.%</i>	<i>Sens.%</i>	<i>Spec.%</i>
APC promoter hypermethylation	93.4	95.4	94.2	95.9	94.4	95.9	92	94.4
<i>RARβ_2 promoter hypermethylation</i>	95.5	92.4	95.3	92.4	95.5	92.4	90	100
CEA	52.1	76.7	55.8	76.6	53.9	76.6	58	74.9
CA15.3	75.2	86.2	76.7	86.2	74.2	86.2	76	74.6

Thank You

